Finding Patterns of Opioid Addiction in the Brain's Reward Pathway Esha Dhawan, Lambert High School, Suwanee, GA, 30024 & BioScience Project, Wakefield, MA, 01880

Background

- Opioids work by attaching to specific proteins called opioid receptors, found in the brain and spinal cord in addition to other organs of the body. The euphoric response to opioid medications is a result of an effect on brain regions involved in reward.
- Functional imaging studies have shown that activated regions of the brain during drug intoxication include the nucleus accumbens (NAc), ventral tegmental area (VTA), and frontal lobe (FL).
- The VTA is a major dopaminergic area of the brain that works closely with the nucleus accumbens, which includes important brain circuits involved in reward. The frontal lobe may be damaged during addiction, resulting in a lack of cognitive control and increased impulsivity.

Methods

- The Allen Brain Atlas (http://www.brain-map.org) is a database used to collect gene expression data for the chosen brain regions in comparison with the gray matter of the brain using a differential search. Data for the heat maps was collected from six available donors: H0351.1009, H0351.1012, H0351.2001, H0351.2002, H0351.1015, and H0351.1016. Data for the remaining results was collected from four donors: H0351.1009, H0351.1012, H0351.2001, and H0351.2002.
- Venny 2.1.0 (<u>http://bioinfogp.cnb.csic.es/tools/venny/</u>) was used to compare gene lists from four chosen brain donors to identify genes that are common and different in each. A Venn diagram is created as a visual representation of the data.
- Python Anywhere (<u>https://www.pythonanywhere.com</u>) is a programming tool used to calculate the statistic values and variance of the fold change values of each of the four gene lists and create histograms as a visual representation.
- DAVID (https://david.ncifcrf.gov) is a bioinformatics clustering tool that subdivided the gene lists based on varying criteria related to function. Gene lists were sorted using the "official gene symbol" identifier and limited to annotations of "homo sapiens". Functional annotation tools were used to analyze the results.
- Genes of interest were entered in the STRING database (<u>http://string-db.org</u>) to identify potential interacting partners, pathways, and other genes relating to addiction. The database consists of networks with experimentally validated interactions.
- Genes of interest were also entered in GeneWeaver (<u>http://www.geneweaver.org</u>) to provide further information about the relevant genes and their functions by searching numerous experimental databases.

Results

Gene Expression Profiles

Ventral Tegmental Area

Nucleus Accumbens

Frontal Lobe



The heat maps demonstrate microarray data showing gene expression profiles of six donors (H0351.1009 H0351.1012, H0351.2001, H0351.2002, H0351.1015, H0351.1016). Each column represents a tissue sample. This data is collected from mRNA that is copied into cDNA and labeled and hybridized to an array containing all human genes. Data with a fold change of ~ 3 or above was used in the analysis.

Two different sample types are used for comparison: the sample under study and the control. The heat maps range in color based on the z-score over a probe. Red areas of the heat maps indicate that the expression of the sample is greater than the control (z-score of +3 and above), green areas show that the expression is less than the control (z-score of -3 and below), and black areas show that the expression is equal to the control (z-score of 0).

2. Gene Overlaps Among Chosen Donors in Each Region

Ventral Tegmental Area



87 genes (8.5%) common in all donors



242 genes (26.7%) common in all donors Frontal Lobe



74 genes (38.7%) common in all donors

3. Top 20 Genes with Highest Fold Change Valu

These Venn diagrams demonstrate the number and percentage of genes that overlap in each region of the brain in four chosen donors (H0351.1009; List 1.

H0351.1012; List 2,

H0351.2001; List 3, H0351.2002; List 4).





The graphs of the genes from the donors shown above display the 20 genes with the top fold change values in the VTA. 19 out of 20 of these genes were found to be common in all four chosen donors.

SLC18A2, DDC, SLC6A3, EN1, and TH were all genes that consisten had the highest fold changes and were found in all four of the chosen donors.





In the nucleus accumbens, 20 out of 20 of the genes with the top fold change values in all four of the donors (including H0351.1009 and H0351.1012) were common in all donors, two of which are shown ab

SAG, PENK, PDYN, and SYNGIG1L consistently had the highest fold change values and were common in all four donors.



4.



In the two donors of the frontal lobe shown above, 20 out of 20 of the genes with the top fold change values were common in all of the donor

TMEM15S, FOXG1, CCK, and LY86-AS1 were all genes that consister had the highest fold change values and were common in all four donor

Statistics and Variance of Genes

Ventral Tegmental Area								
tats	fold-change 1	fold-change 2	fold-change 3	fold-change 4				
ount	276.000000	276.000000	276.000000	276.000000				
ean	7.380185	9.970536	5.865236	8.955087				
td	10.023900	32.448725	4.580648	23.333101				
in	3.633000	2.993000	3.363000	3.574000				
5%	4.095500	3.298250	3.689250	3.896500				
0 %	4.790500	3.860000	4.288500	4.461500				
5%	6.284750	5.596250	5.820000	5.927250				
ax	108.695000	378.537000	41.387000	275.509000				
variance fold-change_1 100.478580 fold-change_2 1052.919780 fold-change_3 20.982337 fold-change_4 544.433621								



Fold-change_1 represents Donor H0351.2001, Fold-change_2 represents [H0351.2002, Fold-change_3 represents Donor H0351.1009, Fold-change represents Donor H0351.1012

In the VTA, the frequency of genes (y-axis) is heavily skewed for gene with a smaller fold change (x-axis). The mean fold change for each do varies slightly, ranging from about 5.9-10. The standard deviation, how varies drastically among donors, the lowest fold change being about 4 and the highest being 32.4.

In the NAc, the distribution of fold changes was similarly represented mean fold change was approximately the same in each donor, each about 7. The standard deviation in each donor was also approximately the same, each about 8.

	35				
stats					30 -
count	fold-change_1 101.000000 3.872386	fold-change_2 101.000000 5.113317	fold-change_3 101.000000 3.909030	fold-change_4 101.000000 4.405614	25 -
std min	0.918703 2.918000	1.670388	1.192413 2.963000	1.226790	20 -
25% 50%	3.178000 3.561000	3.934000 4.521000	3.142000 3.462000	3.593000 3.986000	15 -
75% max	4.263000 6.576000	5.400000 10.639000	4.016000 8.500000	4.656000 9.230000	10 -
variance fold-change 1			0.844015		5 -
	f f f	old-change_2 old-change_3 old-change_4	2.790197 1.421849 1.505014		0 2.5 3
	•				Histo

fold-change_1

Histogram of Donor H0351.2001

The fold-change columns represent the same donors as above.

In the frontal lobe, the frequency of genes is less skewed. However, overall, the fold change values are smaller than those in the VTA and NAc. The maximum fold change value in the frontal lobe was 10.6.

The mean fold change is relatively similar in all donors, ranging from 3.9-5.1. The standard deviation is also nearly the same, ranging from 0.9–1.7.

Jes	5.	Gen	es of Interest				
6 6		Annotation Cluster 2 Enrichment Score: 3 C GOTERM_BP_FAT catecholomine biosynthetic process RT GOTERM_BP_FAT catecholomine metabolic process RT GOTERM_BP_FAT colourine biosynthetic process RT GOTERM_BP_FAT docamine biosynthetic process RT GOTERM_BP_FAT biogenic amine biosynthetic process RT GOTERM_BP_FAT biogenic amine biosynthetic process RT GOTERM_BP_FAT biogenic amine biosynthetic process RT GOTERM_BP_FAT calubal amino acid derivative metabolic process RT GOTERM_BP_FAT calubal amino acid derivative metabolic process RT GOTERM_BP_FAT calubal amino acid derivative metabolic process RT GOTERM_BP_FAT atlobol bindins					
ntly	Results from the Ventral Tegmental Area						
		GOTERM, BP_FAT boomstary, behavior RT i i MITERMAO Oopamine resource RT i i GOTERM, BP_FAT dopamine resource available, process RT i i GOTERM, BP_FAT contransmitter, uptake RT i i GOTERM, BP_FAT contrainsmitter, uptake RT i i GOTERM, BP_FAT containsmitter, uptake RT i i GOTERM, BP_FAT containsmitter, uptake RT i i GOTERM, BP_FAT creation frithrith RT i i GOTERM, BP_FAT creation frithrith RT i i GOTERM, BP_FAT creation frithrith RT i i GOTERM, BP_FAT resource to organic cyclic subtater RT i i GOTERM, BP_FAT resource to organic cyclic subtater RT i i GOTERM, BP_FAT resource to organic cyclic subtater RT i i GOTERM, BP_FAT </th <th>1 1.0 2.0 1.0 2.0 2.0 0.0</th>	1 1.0 2.0 1.0 2.0 2.0 0.0				
			6 5.8E4 3.1E1				
ove. d		NOTERM_BP_FAT cell morphogenesis involved in neuron RT NOTERM_BP_FAT neuron projection morphogenesis RT NOTERM_BP_FAT neuron development RT NOTERM_BP_FAT cell mothogenesis RT NOTERM_BP_FAT cell mothogenesis RT NOTERM_BP_FAT cell mothogenesis RT NOTERM_BP_FAT cell mothogenesis RT NOTERM_BP_FAT cell ant morphogenesis RT NOTERM_BP_FAT cell and comphogenesis RT NOTERM_BP_FAT cell morphogenesis RT <	68.468.4669.169.1678.269.2689.260.00089.260.00069.269.2679.260.00079.260.00079.260.00080.0000.00080.0000.0009 <t< th=""></t<>				
		Resu	Its from the Frontal Lobe				
	An ana numbe genes genes	alysis of the genes commo or of relevant genes of inte of interest, while terms hig of interest.	on of all four chosen donors revealed a large crest. Terms highlighted in green represent general ghlighted in blue represent addiction-related				
rs. ntly rs.	Many of the genes of interest included dopamine-related processes and binding, behavioral responses, neurological system processes, neurotransmitters, and responses to drugs (nicotine, cocaine, alcohol).						
	Relevant genes generating addiction-related responses in all three regions of the brain included CHAT, CHRNA3, CHRNA4, DRD1, DRD2, HTR2C, SLC6A3, OPRD1, OPRM1, and PPP1R1B.						
	6.	Protein Interac	tion Networks and Results				
	Netw	ork found between DRD2 SLC6A3	and Network found between OPRM1, OPRD1, and PENK				
Donor e_4	DRD2 : activity which -gene the VT	dopamine receptor D2; is mediated by G protein nhibit adenylyl cyclase common in all donors in b A and NAc	OPRM1: opioid receptor, mu 1sOPRD1: opioid receptor, delta 1; inhibits neurotransmitter release by reducing calcium ion currents and increasing potassium ion conductance				
onor wever, 1.5	SLC6/ (neuro dopam action	3: solute carrier family 6 transmitter transporter, ine), member 3; terminate of dopamine	PENK: proenkephalin; Met- and Leu- enkephalins compete with and mimic the effects of opiate drugs; play a role in pain perception and responses to stress				
The	Α.	GNAZ	B. COPS5 SIAH1				



ADORA2A:

-gene found to have a response to amphetamine and to be involved in the dopaminergic pathway

adenosine A2a receptor; activity of this receptor is mediated by G proteins which activate adenylyl cyclase

guanine nucleotide binding protein (G protein), alpha activating activity polypeptide O; G proteins are involved as modulators or transducers in various transmembrane signaling systems

KCNF1

GNAO1:

Dok

-gene found to have a response to

morphine and drug

OPRD1

A. The DRD2 network consists of 25 proteins, 30 edges, and has low interconnectivity. Fifteen of these have been implicated in neuro-related processes - GIPC1, NSF, ADORA2A, CALM1, CALM2A, NCS1, GRIA2, GNA12, GNA13, SLC6A3, SSTR5, EPB41, EPB41L1, FLNA, GNAZ. These processes include dopamine signaling, opioid response, and neuronal signaling. Four of these proteins (GRIA2, GNA12, GNA13 and GNAZ: KEGG Pathway) are also associated with long-term depression.

B. The OPRM1 network consists of 28 proteins, 53 edges, and has moderate interconnectivity. Ten of these proteins function in similar neuronal processes as the DRD2 network - GNAO1, GNAI1, GNAI2, OPRM1, PENK, UBC, SIAH1, SIAH2, CALM1, and UBC.

The two networks have 3 proteins in common - CALM1, FLNA, and GNAI2.



Out of the 7 genes that were found in GeneWeaver genesets and KEGG pathways of cocaine, morphine, and amphetamine addiction, **DRD1** was a gene of interest. No genes of interest were found in the genesets of nicotine, cocaine, and amphetamine addiction.

Out of the 36 genes found in the cocaine and amphetamine addiction KEGG pathway genesets, **DDC**, **TH**, **PDYN**, **AND SLC18A2** were genes that were consistently found to have the highest fold-change values in all four donors in the VTA and NAc.



TH and PDYN can be seen in this visual representation of a cocaine addiction KEGG pathway.

Conclusion

- An informatics approach was used for profiling gene expression patterns to identify other genes that may be involved in opioid addiction.
- In the VTA, NAc, and frontal lobe, the genes with the highest fold change values tended to be common in all four chosen donors.
- The genes of interest with the highest fold change values include SLC18A2, DDC, SLC6A3, TH, PENK, and PDYN.
- There is a high conservation of gene expression patterns pertaining to addiction in all three brain regions relevant to the reward pathway, especially in the VTA and NAc.
- Gene interaction networks were found between DRD2, SLC6A3, and ADORA2A, as well as between OPRM1, OPRD1, PENK, and GNAO1.
- The DRD2 and OPRM1 networks both have genes that overlap in KEGG pathways for long-term depression, indicating possible comorbidity between substance abuse and depression. Both networks also indicate a high level of G-protein signaling.
- DRD1 is a dopamine receptor that is activated in morphine, cocaine, and amphetamine addiction.
- GNAO1 and OPRM1 are genes activated only in morphine addiction and therefore may be specific to opioid addiction.